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# IPKine<sup>™</sup> Anti-GFP Magnetic IP Kit

Cat #: KTI2024

Size: 20 T/100 T

| [ <u>;</u> ] | Anti-GFP Magnetic IP Kit   |     |                               |  |  |  |
|--------------|--|-----|-------------------------------|--|--|--|
| REF          | <b>Cat #</b> : KTI2024   | LOT | Lot #: Refer to product label |  |  |  |
|              | <b>Capacity:</b> ≥ 0.6 mg GFP-Tag protein/mL<br>Magnetic Beads   |     | Beads Concentration: 20 mg/mL |  |  |  |
|              | Applications: IP   |     |                               |  |  |  |
|              | Reactivity: Mammals, Bacteria  |     |                               |  |  |  |
| Ĵ.           | Storage: Store according to the recommended storage conditions of each component, stable for 12 months |     |                               |  |  |  |

## **Assay Principle**

Anti-Green Fluorescent Protein-Tag (GFP-Tag) Magnetic Beads are prepared by covalently coupling Anti-GFP-Tag Mouse Monoclonal Antibody to crosslinked Magnetic Beads, useful for detection and capture of fusion proteins containing a GFP-Tag peptide sequence by commonly used immunoprecipitation procedures. The Optimized Anti-GFP Magnetic Beads have more efficient antigen binding capacity. According to the structure, biological function and subsequent application requirements of the target protein, this kit provides two elution methods, including acid elution and denatured elution. Especially by using acid elution will not contain heavy chain and light chain of antibody, which could effectively avoid the interference of heavy chain and light chain of antibody in Western Blotting experiment after immunoprecipitation.

## **Materials Supplied and Storage Conditions**

|             |                              | Size   |        |                    |
|-------------|------------------------------|--------|--------|--------------------|
|             | Kit components               | 20 T   | 100 T  | Storage conditions |
|             | Non-Denaturing Lysis Buffer  | 20 mL  | 100 mL | 4°C                |
|             | TBS (10×)                    | 20 mL  | 100 mL | 4°C                |
|             | Anti-GFP Magnetic Beads      | 0.4 mL | 2 mL   | 4°C, Avoid freeze  |
| Part 1 of 2 | Mouse IgG Magnetic Beads     | 80 µL  | 400 µL | 4°C, Avoid freeze  |
|             | Elution Buffer               | 2 mL   | 10 mL  | 4°C                |
|             | Neutralization Buffer        | 0.2 mL | 1 mL   | 4°C                |
| Part 2 of 2 | SDS-PAGE Loading Buffer (5×) | 1 mL   | 2 mL×2 | -20°C              |

# **Materials Required but Not Supplied**

- Magnetic Separation Rack
- Vertical rotating mixer



- Freezing Centrifuge
- Precision Pipettes, Disposable Pipette Tips
- Deionized Water
- PBS Buffer
- Dounce homogenizer (for tissues)

## **Reagent Preparation**

**Non-Denaturing Lysis Buffer:** Native protein lysis buffer, extract protein for IP samples. Ready to use as supplied. Place it on ice for use. Store at 4°C.

1×TBS: Add Deionized Water to the 10×TBS and dilute the 10×TBS to 1×TBS before use. Store at 4°C.

Anti-GFP Magnetic Beads: Ready to use as supplied. Store at 4°C, Avoid frozen.

Mouse IgG Magnetic Beads: Ready to use as supplied. Store at 4°C, Avoid frozen.

Elution Buffer: Ready to use as supplied. Store at 4°C. Used for acid elution of non-denatured proteins.

Neutralization Buffer: Ready to use as supplied. Store at 4°C. Used for neutralize acid elution of non-denatured proteins.

SDS-PAGE Loading Buffer (5×): Ready to use as supplied. Store at -20°C.

Note: (1) Protease inhibitors are not necessarily added, it is recommended that different types of protease inhibitors should be added to the Non-Denaturing Lysis Buffer according to the experimental requirements; (2) It is recommended to use the centrifugal tube with low adsorption for experiment, which can reduce the adhesion of magnetic beads to the centrifugal tube wall. Adding 0.01%-0.1%(V/V) non-ionic detergent (such as Triton X-100, Tween-20 or NP-40) to 1×TBS can also effectively reduce the adhesion of centrifugal tubes to magnetic beads.

### **Assay Procedure**

#### A. Preparation of protein samples

Note: Prepare a certain amount of sample proteins, which were used as whole cell lysate (WCL) for subsequent Western Blotting detection.

#### 1. Extract protein for Cell Samples:

(1) Collect cells (Adherent cells: 80% to 90% of monolayer cells were grown in a 10 cm cell culture dish. Remove the medium and wash with PBS once; Suspended cells: Collect 5×10<sup>6</sup> cells by centrifugation and washed by PBS once.

(2) Add 0.5-1 mL ice-cold Non-Denaturing Lysis Buffer to cells, lytic cells at 4°C for 5 min. During the process, the pipette is used to blow the mixture repeatedly, transfer cell suspension to a new tube.

(3) Centrifuge at 12,000 rpm for 10 min at 4°C, collect the supernatant.

#### 2. Extract protein for Tissue Samples:

(1) Tissue Samples: Weigh 0.1 g of tissue and add 1 mL Non-Denaturing Lysis Buffer, Homogenize tissue with Dounce homogenizer. (If the protein concentration is low, reduce the volume of Non-Denaturing Lysis Buffer).

- (2) Transfer the homogenate to a new tube, lytic samples at 4°C for 5 min.
- (3) Centrifuge at 12,000 rpm for 10 min at 4°C, collect the supernatant.

#### 3. Extract protein for Bacteria Samples:

(1) Collect bacteria by centrifugation (1,2000 rpm for 2 min at 4°C) and washed by PBS once.

(2) Per mL bacteria add 100-200 µL Non-Denaturing Lysis Buffer, Ultrasonic break in ice 5 min (power 20% or 200 W, ultrasonic 3 s, interval 7 s, repeat 30 times).

(3) Centrifuge at 12,000 rpm for 10 min at 4°C, collect the supernatant.

Note: (1) The sample must contain GFP-tag protein and its complex; (2) For Immunoprecipitation, fresh samples are preferred; (3) In Immunoprecipitation experiments, the affinity between different antigen and antibody is different, and the binding of antigen to antibody is also affected by lysis buffer and wash buffer. If the Non-Denaturing Lysis Buffer does not provide the best experimental results, it is recommended to optimize the operation details or screen and prepare suitable by lysis buffer and wash buffer for experiment

**B.** Preparation of magnetic beads

# Note: Per 500 µL of protein sample add 20 µL Anti-GFP Magnetic Beads. Perform the following procedures, according to add 20 µL Anti-GFP Magnetic Beads.

(1) Add Anti-GFP Magnetic Beads to a 1.5 mL centrifuge tube. Place the centrifuge tube on a Magnetic Separation Rack, let stand for 10 s, remove the supernatant.

(2) Add 1 mL 1×TBS to re-suspend Anti-GFP Magnetic Beads, place the tube on a Magnetic Separation Rack, let stand for 10 s, remove the supernatant, repeat 3 times. Add 20 µL 1×TBS to re-suspend Anti-GFP Magnetic Beads.

#### C. Immunoprecipitation

(1) Add 500  $\mu$ L protein samples to the processed Anti-GFP Magnetic Beads, and incubate at room temperature for 1-2 h or overnight at 4°C (It is recommended to use vertical rotating mixer with Low-speed rotation).

Note: a) It is recommended to add Mouse IgG Magnetic Beads for immunoprecipitation in some samples as negative control, which could exclude the non-specific binding of IgG to the target protein or other specific biological molecules; b) In the case of very high background after immunoprecipitation using Anti-GFP Magnetic Beads, it is recommended that Mouse IgG Magnetic Beads be used to preprocess samples to eliminate non-specific adsorption, then use Anti-GFP Magnetic Beads for immunoprecipitation.

(2) Place the tube on a Magnetic Separation Rack, let stand for 10 s, remove the supernatant. The supernatant can be transferred to a new centrifuge tube to test the effect of immunoprecipitation.

(3) Add 1 mL 1×TBS, and re-suspend Anti-GFP Magnetic Beads, place the tube on a Magnetic Separation Rack, let stand for 10 s, remove the supernatant, repeat 3-5 times, until OD280 of the supernatant is lesser than 0.05.

(4) Elution

a) **Denatured elution:** This method is suitable for SDS-PAGE and Western Blotting analysis of elution samples. Add 100  $\mu$ L (5 times volume of Beads) 1×SDS-PAGE Loading Buffer (Dilute the SDS-page Loading Buffer (5×) by 5 times with 1×TBS) to the tube and mix well, incubate at 100°C for 5 min, then Centrifuge at 800 rpm for 1 min, and collect the supernatant to a new tube for SDS-PAGE and Western Blotting analysis.

b) Acid elution: This method can maintain their original biological activity, elution can be used for functional analysis. Add 100  $\mu$ L (5 times volume of Beads) Elution Buffer to the tube and mix well, incubate at room temperature for 5-10 min (It is recommended to use vertical rotating mixer with Low-speed rotation), then centrifuge at 800 rpm for 2 min at 4°C, and collect the supernatant which is GFP-Tag protein and its complex to a new tube, and immediately add 10  $\mu$ L Neutralization Buffer to adjust the pH to 7.0-8.0. In order to improve the elution efficiency, elution can be repeated, and combine the same samples. Place GFP-Tag protein and its complex on ice to be used, or store at -20°C/-80°C for long-term. It is recommended to add 100  $\mu$ L 1 × SDS-PAGE Loading Buffer to beads precipitation to test the effect of immunoprecipitation and elution.

Note: a) For a few samples, due to differences in target proteins, the binding of GFP-Tag and Anti-GFP antibody is very strong, and the effect of Acid elution may be poor. Therefore, SDS-PAGE Loading Buffer denaturation elution method is recommended as a priority; b) Due to the difference of target protein, the elution efficiency of acid elution method also varies to some extent. If the requirement of elution efficiency is high, the pH value of acidic eluent can be adjusted appropriately between 2.5-3.1, and the pH value or quantity of corresponding neutralizing solution should be adjusted appropriately. For example, 100  $\mu$ L Acid Elution Buffer (0.1 M Glycine-HCI, pH 2.8) and 15  $\mu$ L Neutralizing Buffer (1 M Tris-HCI, pH 8.5).

# **Typical Data**

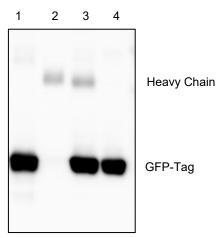


Figure. The immunoprecipitation effect of Anti-GFP Magnetic IP Kit used for GFP-Tag fusion protein.

Data provided for demonstration purposes only. 3/5

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HEK293T cells were transfected with GFP-Tag plasmid, after 48 h, cells were lysed using Non-Denaturing Lysis Buffer after transfection, and then the sample was used for immunoprecipitation, Western Blotting primary antibody was Anti-GFP Tag Mouse Monoclonal Antibody (3D3) (Cat #: ABT2020), dilution factor is 1:5000; Secondary antibody was IPKine<sup>™</sup> HRP, Goat Anti-Mouse IgG LCS (Cat #: A25112), dilution factor is 1:2000; SuperKine<sup>™</sup> West Femto Maximum Sensitivity Substrate (Cat #: BMU102-EN) was used for development. Lane 1 was whole cell lysate (WCL); Lane 2 was the immunoprecipitation sample of Mouse IgG Magnetic Beads eluted by 1×SDS-PAGE Loading Buffer; Lane 3 was the immunoprecipitation sample of Anti-GFP Magnetic Beads eluted by 1×SDS-PAGE Loading Buffer. Lane 4 was the immunoprecipitation sample of Anti-GFP Magnetic Beads eluted by 1×SDS-PAGE Loading Buffer. Lane 4 was the immunoprecipitation sample of Anti-GFP Magnetic Beads eluted by 1×SDS-PAGE Loading Buffer. Lane 4 was the immunoprecipitation sample of Anti-GFP Magnetic Beads eluted by 1×SDS-PAGE Loading Buffer. Lane 4 was the immunoprecipitation sample of Anti-GFP Magnetic Beads eluted by 1×SDS-PAGE Loading Buffer. Lane 4 was the immunoprecipitation sample of Anti-GFP Magnetic Beads eluted by Elution Buffer. By using acid elution, only contained GFP-Tag fusion protein, did not contain heavy and light chains of antibody.

# FAQ

| Problem              | Cause                                  | Suggested Solution   |  |
|----------------------|--|--|--|
|                      | Protein is not completely eluted       | Change elution methods   |  |
|                      |  | Make sure the protein of interest contains the tagged protein by |  |
|                      | No target protein expressed            | Western blotting or dot blotting analyses                        |  |
|                      |  | 1. Use larger volume of cell lysate                              |  |
|                      | Very low protein expression level      | 2. Optimize expression conditions to raise the protein           |  |
|                      |  | expression level   |  |
|                      | Washes are too stringent               | Reduce the time and number of washes                             |  |
| Very few or no       | Incubation times are inadequate        | Increase the incubation time                                     |  |
| tagged protein       | Interfering substance is present in    | Lysates containing high concentration of DTT,                    |  |
| exists in the eluate |  | 2-mercaptoethanol, or other reducing agents may destroy          |  |
|                      | sample                                 | antibody function, and must be avoided                           |  |
|                      |  | If Western blotting detection is used:                           |  |
|                      |  | 1. Check primary and secondary antibodies using proper           |  |
|                      |  | controls to confirm binding and reactivity                       |  |
|                      | Detection system is inadequate         | 2. Verify that the transfer was adequate by using prestained     |  |
|                      |  | protein marker or staining the membrane with Ponceau S           |  |
|                      |  | 3. Use fresh detection substrate or try a different detection    |  |
|                      |  | system   |  |
|                      | Proteins bind nonspecifically to       | 1. Pre-clear lysate with Mouse IgG Magnetic Beads to remove      |  |
|                      | the monoclonal antibody, insufficient  | nonspecific binding proteins                                     |  |
|                      | washing on magnetic beads, or the      | 2. After suspending beads for the final wash, transfer entire    |  |
|                      | microcentrifuge tubes                  | sample to a clean microcentrifuge tube before Magnetic           |  |
|                      |  | separation   |  |
|                      |  | 1. Increase the number of washes                                 |  |
| Multiple protein     |  | 2. Prolong duration of the washes, incubating each wash for at   |  |
| bands found in       |  | least 15 min   |  |
| the eluate           |  | 3. Choose other wash buffers. Increase the salt and/or           |  |
|                      |  | detergent concentrations in the wash solutions                   |  |
|                      |  | 4. Centrifuge at lower speed to avoid nonspecific trapping of    |  |
|                      |  | denatured proteins   |  |
|                      | The protein is not stable at room      | Immunoprecipitation of the target protein at lower temperature   |  |
|                      | temperature                            | such as 4°C  |  |
|                      | Protein degradation due to             | Add protease inhibitors to cell lysate                           |  |
|                      | proteases activity during purification |  |  |



|  | process |  |
|--|---------|--|
|--|---------|--|

## **Recommended Products**

| Catalog No. | Product Name   |
|-------------|--|
| KTD104-EN   | Universal IP/Co-IP Toolkit (Magnetic Beads)                            |
| ABT2024     | Magnetic Beads Conjugated Anti-GFP Tag Mouse Monoclonal Antibody (3D3) |

# Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.

